## Prime Editing precisely corrects prevalent mutations observed in Glycogen Storage Disease Type 1b (GSD1b) patients

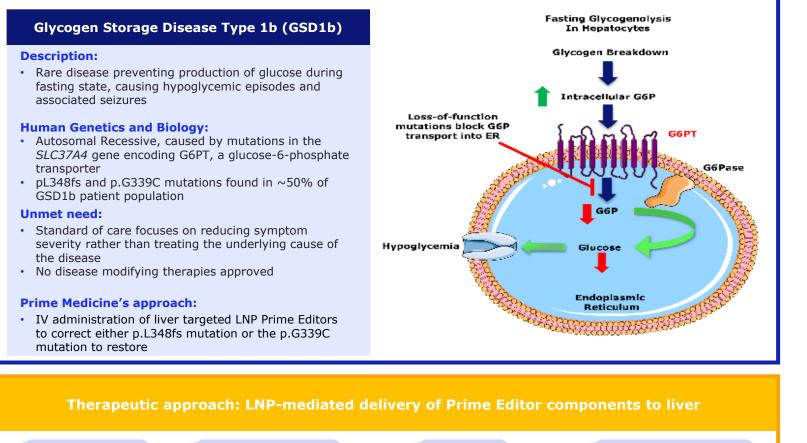
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## Abstract

Glycogen Storage Disease Type 1b (GSD1b) is an autosomal recessive disorder caused by mutations in the SLC37A4 gene encoding the glucose 6-phosphate translocase (G6PT) which is required for normal glucose-6-phosphate metabolism, including hepatic glycogenolysis. Patients exhibit multiple clinical manifestations including severe hypoglycemia resulting in seizures and cognitive impairment. Without an approved treatment for GSD1b, patients maintain metabolic control with a special diet and with medications that alleviate secondary complications such as neutropenia. The most prevalent mutations include p.G339C and/or p.L348fs, observed in ~50% of patients. A gene editing approach to correct the mutations in the affected cells to restore G6PT function would directly address the underlying genetic cause of the disease

Prime Editing (PE) is a next generation gene editing technology that can precisely correct more than 90% of all pathogenic human mutations without the need for double strand breaks (DSBs), with the potential for minimal byproducts at the edit site, and low off-target activity and chromosomal alterations or genotoxicity sometimes observed with CRISPR-based editing. We have developed a universal liver-targeted lipid nanoparticle (LNP) for delivery of Prime Editors to the liver for multiple indications. LNP-RNA PE candidates, that are LNPs formulated with an engineered mRNA encoding the Prime Editor protein and the Prime Editor guide RNA (pegRNA), were developed to correct the mutated SLC37A4 gene in hepatocytes.

Comprehensive high-throughput screening for pegRNAs identified initial hits that correct either the p.G339C or p.L348fs mutation. Initial PE lead assessment in primary hepatocytes isolated from humanized mice in which the mouse Slc37a4 gene was replaced with the human SLC37A4 gene harboring either the G339C or L348fs mutations, or in iPSC-derived hepatocytes, resulted in editing efficiencies up to 80% in vitro. A similar assessment was performed in vivo following intravenous delivery of the LNP-RNA PE candidates to humanized mice. Genomic correction of the L348fs mutation was observed in whole liver at an average correction rate of 47% (total liver alleles) and an associated correction of SLC37A4 transcripts, transcript levels and protein expression. Intravenous administration of LNP-RNA to cynomolgus monkey was well tolerated and resulted in precise editing rates in whole liver of up to 50% at day 14 in animals without significant on-target unintended edits detected. These results demonstrate efficient and safe LNPmediated delivery of Prime Editor candidates to the liver and that Prime Editing can efficiently and precisely correct pathogenic mutations causing GSD1b at rates exceeding those believed to reverse manifestations of disease.



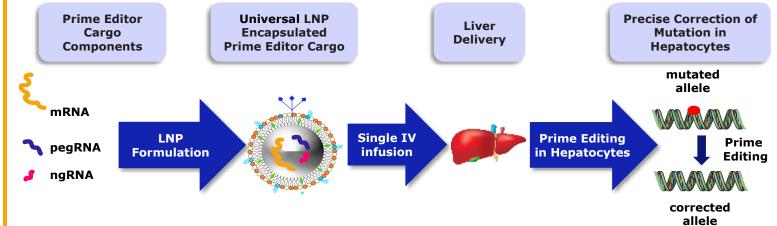
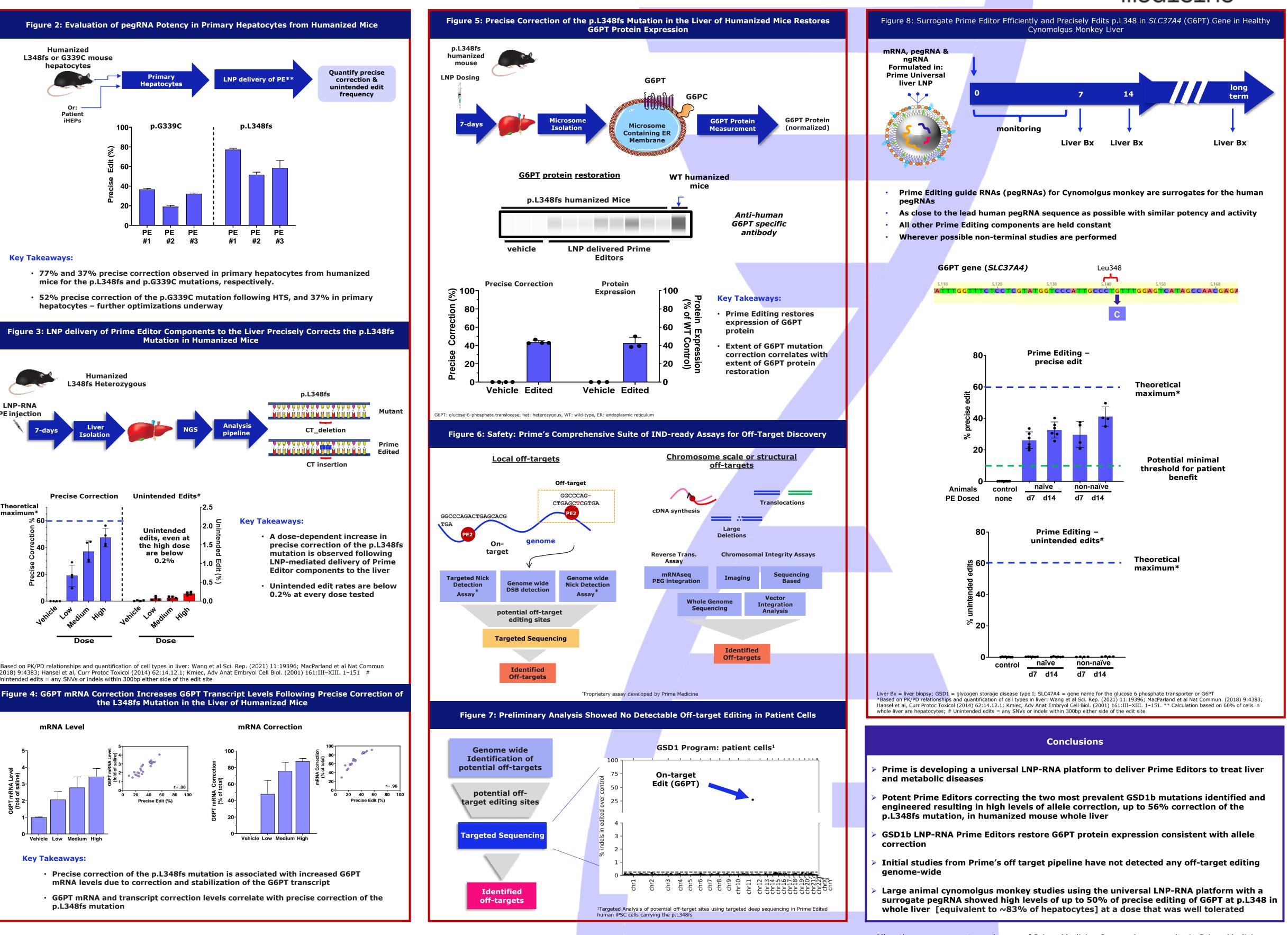


Figure 1: Guide Screening Pipeline Identified Prime Editors That Precisely Correct the G339C and L348fs Mutations p.G339C p.L348fs Precise Correction **Precise Correction** 100 Unintended Edit Unintended Edits<sup>#</sup> All Data **Top Hits** All Data Top Hits **Key Takeaways:**  Guide screening pipeline identified highly active candidates that can precisely correct the p.G339C and p.L348fs mutations.

+Unintended edits = any SNVs or indels within 300bp either side of the edit site; \*Data shown using humanized primary mouse hepatocytes; \*\*PE = Prime Editor

Results

Humanized L348fs or G339C mouse hepatocytes Patient iHEPs Key Takeaways: hepatocytes – further optimizations underway Humanized L348fs Heterozygous **LNP-RNA PE injection** Precise Correction Theoretical maximum\* nintended edits = any SNVs or indels within 300bp either side of the edit site **mRNA** Level Vehicle Low Medium Hig **Key Takeaways:** p.L348fs mutation



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